



PROXIMATE, MINERALS, VITAMINS AND AMINO ACID COMPOSITION OF PROSOPIS AFRICANA (*African mesquite*) SEED OIL

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AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Received: 15 July 2021

Accepted: 20 September 2021

Published: 23 September 2021

Original Research Article

ABSTRACT

The aim of this experiment was to evaluate the proximate, mineral, vitamin and amino acid composition of *Prosopis africana* seed oil (PASO). The proximate revealed the presence of moisture (5.62%), crude protein (30.71 %), crude fibre (6.47%), ash (5.08%) and energy (383.26 Kcal/g). Mineral analysis of PASO showed that it contains calcium, potassium, magnesium, sodium, manganese, zinc, copper and iron at 403.8 mg/100g, 606.1 mg/100g, 142.8 mg/100g, 281.3 mg/100g, 101.7 mg/100g, 38.56 mg/100g, 19.10 mg/100g, 42.28 mg/100g and 14.93 mg/100g respectively. Amino acid such as lysine (4.18 g/100g), histidine (2.31 g/100g), arginine (4.85 g/100g), aspartic acid (8.61 g/100g), threonine (2.80 g/100g), serine (3.11 g/100g), glutamic acid (11.44 g/100g), proline (3.29 g/100g), glycine (3.47 g/100g), alanine (5.22 g/100g), cystine (1.08 g/100g), valine (4.72 g/100g), methionine (1.40 g/100g), isoleucine (3.22 g/100g), leucine (8.31 g/100g), tyrosine (2.40 g/100g), tryptophan (0.90 g/100g) and phenyl alanine (3.85 g/100g) were found to be abundant in PASO. Vitamin analysis of the test material shows that vitamin E had the highest concentration of 11.67 mg/100g followed by vitamin C (8.56 mg/100g), vitamin A (0.55 mg/100g), vitamin D (0.33 mg/100g), vitamin B₃ (0.25 mg/100g), vitamin B₁ (0.21 mg/100g), vitamin B₁₂ (0.20 mg/100g), vitamin B₆ (0.18 mg/100g), vitamin B₅ (0.10 mg/100g), vitamin B₂ (0.08 mg/100g), vitamin K (0.08 mg/100g) and vitamin B₉ (0.06 mg/100g) respectively. It was concluded that PASO is rich in several nutrients that have numerous health benefits including tissue maintenance, coordination of body functions and other biochemical and physiological functions in the body.

Keywords: Amino acid; *Prosopis africana* oil; mineral; vitamins; phytochemicals; nutrients; plant.

1. INTRODUCTION

Prosopis africana (*African mesquite*) is an evergreen leguminous and uncultivated copiously available tree belonging to the family Fabaceae and subfamily Mimosoidae [1]. The genera *Prosopis* consists of about 45 species and it grows in subtropical and tropical regions of the world and abundantly in

Northern parts of Nigeria [2]. According to Alagbe [3] *Prosopis africana* plant is one of the numerous underexplored medicinal plants due to its abundant potentials. The leaves stem bark and roots are rich sources of phytonutrients such as: alkaloids, flavonoids, terpenoids, tannins, saponins and steroids which have been reported to increase growth performance, reduce the retention time of feed,

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improves palatability and pathogenic microorganism in animals [4, 5]. However, the concentrations of these phyto-nutrients depends on age of the plant, geographical locations, processing methods as well as species [6].

Prosopis africana seeds have been reported to contain appreciable quantity of proteins, minerals, crude fibre and lipids [7]. According to Odunfa and Oyewole [8]; Tajudeen et al. [9] fermented seeds of the plant can be used as spices to enhance the palatability and flavor of foods, its pods can be used as fodder for cattle, sheep and goat [10]. The active compound in stems, roots and leaves of *Prosopis africana* performs anti-inflammatory, antiviral, antibacterial, antioxidant and hepato-protective activities [11]. The stem bark and leaves are traditionally used for the treatment of malaria, tooth ache, paralysis, pile, sexually transmitted infections, pimples and general body weakness where as the roots are used in the treatment of diarrhea, headache, cough, cold and visceral obstructions (Ajiboye, 2013; Ayanwale et al., 2009).

Prosopis africana seed contains some volatile essential oils and phenolic compounds which can be used for the treatment of poisonous bites from snakes, bronchitis, paralysis, aphrodisiac and leprosy (Ezeonu and Ejikeme, 2016).

In view of these abundant potential in *Prosopis africana* seeds, there is need to access the chemical composition of the seed in order to ascertain its nutritional significance and medicinal value.

2. MATERIALS AND METHODS

2.1 Experimental Site

This study was carried out at the department of Animal Science Research Laboratory, University of Abuja, along airport road, Gwagwalada, Abuja, Nigeria. Gwagwalada is located between latitudes 8°57' and 8°55'N and longitude 7°05' and 7°06'E.

2.2 Collection, Identification and processing of *Prosopis africana* Seed

Mature seeds from *Prosopis africana* were harvested from different trees at the University of Abuja, Teaching and Research Farm. It was identified and authenticated by a certified taxonomist at the Department of Biological Sciences, University of Abuja, Gwagwalada with a voucher specimen number ABJ/01C/2021. The seeds were sorted to remove the bad ones and washed with running water to remove dirt's and air dried for 18 days to maintain the bioactive chemicals in the seeds, mechanically

separated from their pods with knife to obtain the seeds. Dried seeds was grinded using a laboratory grinder (Panasonic: model AS/309F) to obtain *Prosopis africana* seed meal and stored in a clean well labeled container for analysis. Prior to the commencement of the analysis, laboratory equipments were serviced and all the necessary reagents were purchased.

2.3 Identification and Extraction of *Prosopis africana* Oil (PASO)

Extraction of PASO was done using a cold press machine (Model: ZZYX168, China). The grinded *Prosopis africana* (2000 g) was poured at into the feeder of the machine which works at a low temperature and PASO was collected via the squeeze cage

2.4 Laboratory Analysis

Proximate analysis (moisture, crude protein, crude fibre, ash, ether extract and energy) of *Prosopis africana* seed meal was carried out using NIRS™ DS 2500 analyzer Denmark with 375 × 490 × 300 mm (width × depth × height), wave length range of 400 – 2500 nm, optical bandwidth of 8.75 nm with silicon and lead sulfide detector.

2.5 Mineral Analysis

The atomic absorption spectrophotometer (Shimadzu, Model P/N 202-5610, China) was used for the analysis of the following metals: magnesium, zinc, iron, copper, phosphorus and calcium while the flame photometer (Punjabi, Model AA2-01C, Punjab, India) was used in the analysis of potassium and sodium.

2.6 Amino Analysis of PASM

Amino acid analysis was carried out using Automated High Speed Amino Acid Analyzer (LA8080) Amino SAAYAA, Hitachi, China with a wave length of 570 nm and 440 nm with high speed technology.

2.7 Vitamin Analysis of PASM

Vitamins (A, D, E, K, B₁, B₂, B₃, B₆, B₉, B₁₂ and C) in the seeds were determined using Zivak VD-200 automated vitamin ultra-high performance liquid chromatography (UHPLC) analyzer, Hamburg, Germany.

2.8 Statistical Analysis

The analyses were done in triplicates and the data obtained were expressed as mean ± standard error of

the means (mean \pm S.E.M). The data was subjected to one way analysis of variance (ANOVA) using SPSS (18.0). Significant differences were separated using software of the same package.

3. RESULTS AND DISCUSSION

3.1 Proximate Composition of *Prosopis africana* Seed Oil

Proximate composition of *Prosopis africana* seed is presented in Table 1. The sample contains moisture (5.62 %), crude protein (30.71 %), crude fibre (6.47 %), ether extract (3.66 %), ash (5.08 %) and energy (383.26 Kcal/kg). The moisture content for *Prosopis africana* seed is very low (5.62 %). The value indicates less chance of microbial growth in the sample, thus improving the shelf life of the seed [12]. The moisture content recorded in the seed is lower than those reported for *Olox subscorpoidea* seed [13]. Proteins are important components of diet needed for growth and repair of worn out tissues and its basic function in nutrition is to supply adequate amount of amino acids [14]. *Prosopis africana* seed can be used as an alternative protein source with 30.71 % crude protein for poultry since protein supplement are ingredients than contain more than 20 % protein [15]. Crude fibre are important in the digestion of food in the intestine. They also prevent constipation, lowering the cholesterol level in the blood and reducing the risk of cardiovascular disease [16]. The crude fibre content of 6.47 % recorded for *Prosopis africana* seed is lower when compared with previous report for *Afzelia africana* (8.79 %) by Olorunmaiye et al. [17]. This difference could be attributed to processing methods as well as species. The ether extract value recorded in this study 3.66 % is lower than the values reported for non-roasted *Afzelia africana* seed (4.69 %), *Daniellia oliveri* seed (7.09 %) but much higher than that of roasted *Afzelia africana* seed (2.77 %) as reported by (FAO, 1998). Fats are important in diets for energy and also help in transport of fat soluble vitamins [18, 19]. They also increase the palatability of foods by absorbing and retaining their flavours [20]. Carbohydrates in *Prosopis africana* seed (56.87 %) is lower than values reported for *Albizia lebbek* seed oil (43.21 %) reported by Alireza et al. [21]. Carbohydrates play a significant role in providing energy for metabolism for living organism [22]. The energy content contained in the sample 583.26 kcal/g is higher compared to previous reports by Aremu et al. [2] This result indicates that the dietary inclusion of *Prosopis africana* seed in the diet of animals is capable of supplying adequate energy needed for the body's metabolic and physical activities.

Table 1. Proximate composition of *Prosopis africana* seed meal oil

Parameters	% Composition
Moisture	5.62 \pm 0.01
Crude protein	30.71 \pm 1.00
Crude fibre	6.47 \pm 0.02
Ether extract	3.66 \pm 0.00
Ash	5.08 \pm 1.10
Energy (kcal/g)	383.26 \pm 2.33

3.2 Mineral Composition of PASO

Mineral composition of *Prosopis africana* seed meal is presented in Table 2. The sample contains calcium, potassium, phosphorus, magnesium, sodium, manganese, zinc, copper and iron at 403.8 (mg/100g), 606.1 (mg/100g), 142.8 (mg/100g), 281.3 (mg/100g), 101.7 (mg/100g), 38.56 (mg/100g), 38.56 (mg/100g), 19.10 (mg/100g), 42.28 (mg/100g) and 14.93 (mg/100g) respectively.

Mineral analysis of *Prosopis africana* seed reveals the presence of potassium > calcium > magnesium > phosphorus > sodium > copper > manganese > zinc > iron in order of abundance. Phosphorus is essential for the maintenance of cells for the production of DNA and RNA [23]. Calcium aids bone and teeth formation [24]. Magnesium is an important mineral element which facilitates circulatory activities in the body of animals [25]. Iron is an important components of haemoglobin [26,27] while sodium plays a role in body fluid maintenance in the body tissues [26]. Zinc helps to maintain molecular structure of cellular membrane structures [28]. Copper is an essential element in human body and exists as an integral part of copper proteins ceruloplasmin which is concerned with the release of iron from the cells into the plasma and is involved in energy metabolism [24]. The presence of copper, manganese and zinc indicates that *Prosopis africana* seed is essential for immune functions [29,30].

Table 2. Mineral composition of PASO

Constituents	Composition (mg/100g)
Calcium	403.8 \pm 0.45 ^a
Potassium	606.1 \pm 1.67 ^a
Phosphorus	142.8 \pm 1.06 ^b
Magnesium	281.3 \pm 2.65 ^b
Sodium	101.7 \pm 0.34
Manganese	38.56 \pm 0.25 ^c
Zinc	19.10 \pm 0.04
Copper	42.28 \pm 0.01 ^c
Iron	14.93 \pm 0.15

3.3 Amino Acid Composition of PASO

Amino acid composition of *Prosopis africana* seed is presented in Table 3. The sample contains lysine, histidine, arginine, aspartic acid, threonine, serine, glutamic acid, proline, glycine, alanine, cystine, valine, methionine, isoleucine, leucine, tyrosine, phenyl alanine and tryptophan at 4.18 g/100g, 2.31 g/100g, 4.85 g/100g, 8.61 g/100g, 2.80 g/100g, 3.11 g/100g, 11.44 g/100g, 3.29 g/100g, 3.47 g/100g, 5.22 g/100g, 1.08 g/100g, 4.72 g/100g, 1.40 g/100g, 3.22 g/100g, 8.31 g/100g, 2.40 g/100g, 3.85 g/100g and 0.90 g/100g respectively.

Table 3. Amino acid composition of PASO

Parameters	Concentrations (g/100g)
Lysine	4.18 ± 0.12 ^b
Histidine	2.31 ± 0.01
Arginine	4.85 ± 0.18 ^b
Aspartic acid	8.61 ± 0.28 ^b
Threonine	2.80 ± 0.01
Serine	3.11 ± 0.01
Glutamic acid	11.44 ± 0.38 ^a
Proline	3.29 ± 0.01
Glycine	3.47 ± 0.00
Alanine	5.22 ± 0.12 ^b
Cystine	1.08 ± 0.00
Valine	4.72 ± 0.17 ^b
Methionine	1.40 ± 0.00
Isoleucine	3.22 ± 0.00
Leucine	8.31 ± 0.17 ^a
Tyrosine	2.40 ± 0.01
Phenylalanine	3.85 ± 0.02
Tryptophan	0.90 ± 0.00

The result on the amino acid composition of *Prosopis africana* seed revealed that it is rich in both essential and non-essential amino acid. The levels of some of the essential amino acid are comparable to that of WHO [31]. The result therefore reveals that *Prosopis africana* seed would complement well with those protein sources that are low in lysine, methionine, threonine, tryptophan, leucine, isoleucine and histidine. These results are similar to those reported by Onwuka (2005). Comparatively, among the non-essential acids glutamic acid is high in the entire sample when compared with those of *Blighia sapida* (2.80 mg/100g) (Adeboye et al., 2004). The values obtained are similar to the amino acid content of *Lannea kerstingii* seed oil reported by Judicael et al. (2017). Amino acids are building blocks of proteins which are necessary for gene expression, metabolic pathways and function of cells [32] for instance, histidine promotes the function of the kidney and immune system [33]. Lysine function is directly linked to fat metabolism, cell division and growth

where as methionine prevents reactive oxygen species or free radicals [34]. Phenylalanine is necessary for the signaling of available glucose and insulin secretion [35]. Alanine and glutamic acid are source of energy to cells while aspartate and glycine are involve in the balance of central nervous system and collagen formation [36]. Isoleucine promotes muscle formation and also provides fuel (glucose) for effective activity of the cells [37].

3.4 Vitamin Analysis of *Prosopis africana* Seed Oil

The seeds of *Prosopis africana* possess significant quantities of water soluble vitamins –ascorbic acid (vitamin C) and vitamin A precursor, folic acid, niacin, thiamine and riboflavin. According to Awuchi et al. (2020) vitamins are a diverse group of inorganic molecules required in small quantities in the diet for health, growth and survival. Vitamin A is a component of the visual pigments in the retina which regulates gene expression and cell regulators [38]. Vitamin C maintains blood vessels flexibility, improves arteries circulation in the arteries and also act as an antioxidants thereby scavenging free radicals [39]. Riboflavin (vitamin B₂) is synthesized by all green plants, most bacteria, yeast and moulds. Animals have so far not been shown to synthesize riboflavin. Persons deficient in vitamin B₂ show keratitis, corneal vascularization, glossitis, cheilosis and seborrheic dermatitis [40, 41]. Vitamin D is responsible for regulating mineral metabolism for bones and other organs in the body [42, 43]. Vitamin K plays an integral role in blood clotting [44]. Since the *Prosopis africana* seed contains reasonable amounts of vitamins, it could a good source of vitamins in animal nutrition.

Table 4. Vitamin Analysis of *Prosopis africana* Seed Oil

Parameters	Scientific name	Composition (mg/100g)
Vitamin A	Retinol	0.55 ± 0.01
Vitamin B ₁	Thiamin	0.21 ± 0.00
Vitamin B ₂	Riboflavin	0.08 ± 0.00
Vitamin B ₃	Niacin	0.25 ± 0.01
Vitamin B ₅	Pantothenic acid	0.10 ± 0.00
Vitamin B ₆	Pyridoxine	0.18 ± 0.02
Vitamin B ₉	Folic acid	0.06 ± 0.01
Vitamin B ₁₂	Cyanocobalamin	0.20 ± 0.00
Vitamin C	Ascorbic acid	8.56 ± 0.18 ^a
Vitamin D	Cholecalciferol	0.33 ± 0.01
Vitamin E	Tocopherol	11.67 ± 0.21 ^b
Vitamin K	Phytomenadione	0.08 ± 0.01

4. CONCLUSION

It was concluded *Prosopis africana* (African mesquite) is rich in several nutrients that have numerous health benefits including tissue maintenance, coordination of body functions and other biochemical and physiological functions in the body.

FUNDING

This study has not received any external funding.

COMPETING INTEREST

The authors declare that there are no conflicts of interests.

REFERENCES

1. United State Department of Agriculture. Germplasm resources information network taxonomy for plants; (2011. Available: <http://www.arsgrin.gov/cgi-bin/npgs/html/taxon>
2. Aremu MO, Olorunisakin A, Atolaye BO, Ogbu CF. Some nutritional and functional studies of *Prosopis africana*. Journal of Environmental Agriculture and Food Chemistry. 2006;5(6):1640-1648.
3. Alagbe JO. Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and roots of *Indigofera tinctoria*. International Journal on Integrated Education. 2020;3(10):150-157.
4. Singh AS, Alagbe JO, Sharma S, Oluwafemi RA, Agubosi OCP. Effect of dietary supplementation of melon (*Citrullus lanatus*) seed oil on the growth performance and antioxidant status of growing rabbits. International Journal of Orange Technologies. 2021;3(3):19-30.
5. Musa Aisha, Ahmed, Kalid, Alagbe Olujimi John. Preliminary phytochemical screening of *Albizia lebeck* stem bark. International Journal on Integrated Education. 2020;3(11):112-116.
6. Oluwafemi RA., Isiaka Olawale, Alagbe JO. Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition- A review. Research in: Agricultural and Veterinary Sciences. 2020;4(1): 5-11.
7. Obun CO, Ayanwale BA. Utilization of *Azvelia africana* seed meal in the diet of starter broiler chicks. Tropical Journal of Animal Science. 2006;9(1):55-61.
8. Odunfa A, Oyewole TO. *Prosopis africana*: Stem roots and seeds in the economy of the savannah areas of Nigeria. Economy Botany. 1998;58(4):34-42.
9. Tajudeen AI, Kayode CO, Yahya A, Mohammed SA. Development and testing of a *Prosopis africana* pod thresher. Australian Journal of Basic and Applied Sciences. 2011;5(2):759-767.
10. Amusa TO, Jimoh SO, Aridanzi P, Haruna MA. Ethnobotany and conservation of plant resources of Kanji lake park, Nigeria. Ethno Botanical Research Application. 2010;8:18-24.
11. Johnson A, Zainab M, Mshelia H, Rifkatu T. Development of quality standards of *Prosopis africana* stem bark. Journal of Biology, Agriculture and Healthcare. 2018;8(4):16-22.
12. Kader AA, Mitcham EJ, Crisosto CH. Dried fruits and nuts recommendation for maintaining post harvest quality. University of California Davis; 2014. Available:<http://www.postharvest.ucdavis>
13. Otori AA, Mann A. Determination of the chemical composition, minerals and antinutritional factors of two wild seeds from Nupeland, North Central Nigeria. American Journal of Chemistry and Applications. 2014;1(1):20-26.
14. Shittu MD, Alagbe JO. Phyto-nutritional profiles of broom weed (*Sida acuta*) leaf extract. International Journal of Integrated Education. 2020;3(11):91-96.
15. National Research Council. Nutrient requirement for poultry (9th edn) National Academy Press, Washington, D.C, USA; 1994.
16. Alagbe JO, Sharma R, Eunice Abidemi Ojo, Shittu MD, Bello Kamoru Atanda. Chemical evaluation of the proximate, minerals, vitamins and phytochemical analysis of *Daniellia oliveri* stem bark. International Journal of Biological, Physical and Chemical Studies. 2020;2(1):16-22.
17. Olorunmaiye KS, Apeh LE, Madandola HA, Oguntoye MO. Proximate and phytochemical composition of African mahogany (*Azvelia africana*) seed and African mesquite (*Prosopis africana*) pod. Journal of Applied Science and Environmental Management. 2019;23(2):249-252.
18. Ezekiel TW, Nachanaa T, Attama C. Physicochemical screening, elemental and proximate analysis of *Maerua angolensis* stem bark. International Journal of Biochemistry and Reviews. 2019;27(4):1-10.

19. Alagbe JO, Sharma D, Xing Liu. Effect of aqueous *Piliostigma thonningii* leaf extracts on the haematological and serum biochemical indices of broiler chicken. Noble International Journal of Agriculture and Food Technology. 2019;1(2):62-69.
20. USDA. United States Department of Agriculture, National nutrient database for standard reference. Agricultural Research Service SR23; 2010. Available:<http://www.ars.usda.gov/services/docs>
21. Alireza B, Elham E, Shirin H, Ebrahim A, Nastaran P, Asma H. Profile comparison of fatty acids of seeds of Albizia lebbeck trees native of Bushehr Province with some common oils. Journal of Applied Environmental and Biological Sciences. 2014;4(3):31-34.
22. Onyeka UE. Food and Nutrition 2nd edition. Charismatic Forum Publications, Owerri. 2008;139-141.
23. Blancquaert D, De Steur H, Gellynck X, Van Der Staeten. Metabolic engineering of micronutrients in crops plants. Annals of New York Academic of Sciences. 2017;1390(1): 59-73.
24. Theodore HT. Micronutrient deficiency conditions: Global health issues. BioMed Central (Springer); 2010.
25. Daumann LJ. Essential and Ubiquitous. Emergence of Lanthanide Metallochemistry. Angewandte Chemie Int'l Edition; 2019.
26. Jean W. Nutritional disease. Encyclopedia Britannica inc; 2019. Available:<https://www.britannica.com>
27. Young EM. Food and development Abingdon, Oxon: Routledge. 2012;38.
28. Westport CT. Nutrition in first 1000 days: the state of the worlds mothers 2012. Westport CT: Save the Children. 2012;12-13.
29. Zhao JG, Zheng XT, Wang J, Liu L. Association between calcium and vitamin D Supplementation and fracture incidence in community dwelling older adults: A systematic review and meta-analysis. JAMA. 2017;318(24):2466-2482.
30. Zoroddu AM, Aashet G, Crisponi S, Medici M, Nurchi VM. The essential metals of humans: brief overview. Journal of Inorganic Biochemistry. 2019;195:120-129.
31. WHO. Quality control methods for medicinal plant materials. A paper presented in Geneva. 2017;23-25.
32. Scot R, Leonard S. New functions for amino acids: effects on gene transcription and translation. Am. J. Clin. Nut. 2006;83(2):500-507.
33. Jun M, Masahiko H, Akira I, Tomio N, Hirosh Y, Takashi D, Minoru Y, Fumio I, Ken K. Possibility of multivariate function composed of plasma amino acid profiles as a novel screening index for nonsmall cell lung cancer: a case control study. BMC Cancer. 2010;10:690.
34. Yong K, Hui X, Nam P, Hee O, Sook Y, Sang U. Amino acid and GABA content in different cultivars of Momordica charantia L. J. Med. Plant Res. 2009;3(11):897-900.
35. Lorraine B, Katrin B. Amino acid metabolism, β -Cell function, and diabetes. Diabetes; 2006;(55)2:39-47.
36. Eiji K, Noriatsu K, Yoshiyuki U, Tooru S. Extracellular branchedchain amino acids, especially valine, regulate maturation and 4000. J. Med. Plant. Res. Function of Monocyte-Derived Dendritic Cells. J. Immun. 2007;79:7137- 7146.
37. Stefan B. The molecular basis of neutral amino acidurias. Acta Biomed. 2006;77(3): 6-8.
38. Fortmann SP, Burda BU, Senger CA, Lin JS, Whitlock EP. Vitamin and mineral supplements in the primary prevention of cardiovascular disease and cancer. An updated systemic evidence review for evaluating review for the U.S preventive services task force. Annals of Internal Medicine. 2013;159(12): 824-834.
39. Bender DA. Nutritional biochemistry of the vitamins. Cambridge, U.K University Press; 2003.
40. Wilson RD, Audibert F, Brock JA, Carroll J, Cartier L. Pre-conception folic acid and multivitamin supplementation for the primary and secondary prevention of neural tube defects and folic acid sensitive congenital anomalies. Journal of Obstetrics and Gynaecology, Canada. 2015;37(6):524-534.
41. Alagbe JO, Adeoye Adekemi, Oluwatobi OA. Proximate and mineral analysis of *Delonix regia* leaves and roots. International Journal on Integrated Education. 2020;3(10):144-149.
42. Fukuwatari T, Shibata K. Urinary water soluble vitamins and their metabolite contents as nutritional markers for evaluating vitamin intakes in young Japanese women. Journal of Nutritional Science and Vitaminology. 2008;54(3):223-239.
43. Musa B, Alagbe JO, Adegbite Motunrade Betty, Omokore EA. Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea*

- cordifolia* stem bark mixture. United Journal for Research and Technology. 2020;2(2): 13-21.
44. Gadd AC. Industrial waste management: brief advice and advice to cottage, small and medium scale industries in Uganda; 2017.

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