



Effect of Different Seed Treatment on Germination of Fishtail Palm (*Caryota urens*) under Naturally Ventilated Poly-house Conditions

R. J. Makwana ^{a++*}, V. D. Rathva ^{a++} and B.H. Panchal ^{b#}

^a Sheth D. M. Polytechnic in Horticulture Model Farm, AAU, Vadodara 390003, India.

^b Krushi Vigyan Kendra, AAU, Arnej, Gujarat, India.

Authors' contributions

This work was carried out in collaboration among all authors. Author RJM conceived and designed the analysis, collected the data, contributed data or analysis tools, performed the analysis and wrote the paper. Author VDR edited the manuscript. Author BHP wrote the paper. All authors read and approved the final manuscript.

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ABSTRACT

An investigation was conducted to study the effect of different seed treatments comprising of mechanical scarification, chemical and plant growth regulator on germination of Fishtail palm (*Caryota urens*) seeds under naturally ventilated poly-house at sheth D. M. Polytechnic in Horticulture College, Anand Agricultural University, Vadodara during 2021 to 2023. There were total 12 treatments under study which was replicated trice and statistically analysed by completely randomized design at 90 days after sowing. The results revealed that treatment T₁₂ i.e. seeds treated with GA₃ 1000 ppm for one day recorded significantly lesser days to initiation of germination

⁺⁺ Assistant Professor;

[#] Senior Scientist and Head;

*Corresponding author: E-mail: drishimak@aau.in;

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(46.33 days) along with higher germination percentage (86.67 % and 85.00 %), germination rate index (5.90 %/day and 5.70 %/day) and germination index (255 and 243.67), coefficient of velocity of germination (6.18 and 6.02), timpson's germination index (1275 and 1218), modified timpson germination index (42.50 and 40.61) and survival percentage (88.33 and 86.67) at 90 days after sowing during first and second year respectively. While, seed without any treatment kept in open condition treatment T₁ showed poor performance in germination percentage as well as survival percentage. Thus, Fishtail palm (*Caryota urens*) seeds soaked in solution of 1000 ppm GA₃ along with sowing them in polybags and kept under NVPH condition provides better germination and survival.

Keywords: Fishtail palm; seed germination; seed treatment; palm seed germination; palm seedling.

1. INTRODUCTION

Fish tail palm also known as Kithul botanically known as *Caryota urens* L. is a monocotyledon multipurpose tree species belonging to family Arecaceae. This species is mainly distributed in Indian Subcontinent and Southeast Asia where they grow in fields and rainforest clearings. Solitary fishtail palm is a multipurpose tree, cultivated both for its products and as an ornamental specimen. The trunk yields starch (sago), which is edible. Palms are felled to extract the central 'pith', and the sweet-tasting pulp is sundried, powdered and stored and can be used for the preparation of bread. Sap is tapped from the inflorescence, then fermented into an alcoholic drink (palm wine or toddy) or boiled down to make syrup or sugar (jiggery). The stem apex (palm heart or palm cabbage) can be chewed raw or cooked. The leaves produce strong fibers (kitul fiber) that are made into ropes, brushes and baskets.

Slow germination in many species of palms has been attributed to a thick, impervious endocarp and sarcotesta [1,2,3]. Seed scarification using concentrated sulfuric acid has improved germination speed in a few species, but has negative effects on Fishtail palm seeds [4]. The long duration (about 4 months) taken for seed germination is considered as one of the major issues in the propagation of this species [5].

Therefore, an experiment was conducted to explore possibilities of improving the seed germination through mechanical, physical and chemical treatments and to develop a cost effective and rapid seed germination method.

2. MATERIALS AND METHODS

The experiment was conducted during 2022 and 2023 at Sheth D. M. Polytechnic in Horticulture College Research Farm, Anand Agricultural University, Model farm, Vadodara to study the effect of different seed treatment on germination

of fishtail palm. Each treatment of the 12 treatments used involved 60 seeds replicated on 3 batches with completely randomized design. A mature inflorescence which consisted of ripened, partially ripened and matured fruits, was carefully brought down to the ground with the help of a rope controlled by a climber, from a tree of fish tail palm from Horticulture Nursery at B. A. College of Agriculture, AAU, Anand. The collected inflorescences were kept inside a poly bag for 3 days to facilitate ripening of un-ripened fruits. Then the fruits were removed from the inflorescence and collected to a poly bag. Seeds were extracted by pressing the poly bag manually followed by separation of fleshy skin (exo-carp). Extracted seeds were dried under shade for a day. Before every treatment, seeds were soaked in the water, floating seeds were eliminated to remove seeds having damage, porous space and immature growth. Seed having similar morphological characters of seed length 14.5 ± 0.5 mm, seed width 11.0 ± 0.5 mm, seed thickness 7.5 ± 0.5 mm and seed weight 1 ± 0.02 g were selected and 20 seeds distributed for each replication.

There were ten set of treatments viz., seed rubbed with sand paper (T₃), Seed soaking in hot water for 5 min and immediate soak in cold water for 15 min for 2 times (2 days) (T₄), seed soaking in water for 5 days (T₅), Seed soaking in 50 % cow urine for 5 days (T₆), Seed soaking in 100 % cow urine for 5 days (T₇), Seed kept in moist coco peat container wrapped with plastic on top for 5 days (T₈), seed soaked in Profenol 50 % (15 min) for 2 times (2days) (T₉), Seed soaked in GA₃ 1000 ppm for 1 day (T₁₀), seed soaking in Bio NP 10 ml/1 liter water (Azotobactor+PSB) for 2 days (T₁₁) and Seed soaked in HNO₃ 50% for 5 mins (T₁₂). Seeds after the treatments were washed with tap water and sown in the polybags having 20 x 10 cm in size filled with the media comprising of garden soil, sand and vermicompost of 1:1:1 ratio. Another two sets of three replication each were prepared and directly

sown in the polybags and kept in open condition (T_1) as a control and another set along with all other treatments were kept under Naturally Ventilated Poly-house Condition (T_2) for taking observations. Seeds were observed every day up to 90 Days. Numbers of the germinated seeds in each treatment were recorded in 3 day intervals. Germination testing time, capacity and speed of germination were computed as follows.

2.1 Initiation of Germination and Germination Percentage

First day of initiation of germination was calculated from the day seeds were planted and represented in tabular formation. Lower the values indicate a faster initiation of germination in particular treatment [6]. Germination percentage was calculated on the basis of formula given in Rules for Seed Analysis [7].

$$GP = (N/A) \times 100$$

The higher the GP value represents the greater the germination of a seed population [8].

Germination rate index was calculated by following formula

$$GRI = (G1/1) + (G2/2) + \dots + (G30/30)$$

Where, $G1$ =Germination percentage on first day of sowing

$G2$ =Germination percentage on second day of sowing ...

The Germination rate index (GRI) reflects the percentage of germination on each day of the germination period. Higher GRI values indicate higher and faster germination [9].

2.2 Germination Index

Germination index (GI) was calculated on the basis on formula proposed by Goodchild and Walker [10].

$$GI = (30 \times T1) + (29 \times T2) + \dots + (1 \times T30)$$

Where, 1. $T1, T2, \dots, T30$ is number of seed germinated on 1st day, 2nd day ... 30th day.

2. And 30, 29 ... 1 were the weights given to the number of germinated seed on the first to last observation respectively.

In the GI, maximum weight is given to the seeds germinated on the first day and less to those

germinated later on. The lowest weight would be for seeds germinated on the 10th day. Therefore, the GI emphasizes on both the percentage of germination and its speed. A higher GI value denotes a higher percentage and rate of germination [11].

2.3 Coefficient of Velocity of Germination

Coefficient of velocity of germination (CVG) was calculated with the formula proposed by Labouriau [12].

$$CVG = \frac{N1 + N2 + \dots + Nx}{100 \times N1T1 + N2T2 + \dots + NxTx}$$

Where, N =No. of seeds germinated each day and

T = No of days from seeding corresponding to N

The CVG gives an indication of the rapidity of germination. It increases when the number of germinated seeds increases and the time required for germination decreases. Theoretically, the highest CVG possible is 100. This would occur if all seeds germinated on the first day [13].

2.4 Modified Timpson Germination Index

Timson's Germination Index (TGI) [14] is computed as follows,

$$\Sigma k = \sum_{i=1}^k Gi$$

Where, G_i is the cumulative germination percentage in time interval i and k is the total number of time intervals.

With reference to above formula modification was proposed by Khan and Ungar [15], where Timson's index (Σk) is divided by the total time period of germination (T_k)

$$\Sigma k_{mod} = \Sigma k / T_k$$

Two year data for all the observations were collected separately and statistical analysis for individual year was done by the statistical method proposed by Panse and Sukhatme [16].

3. RESULT AND DISCUSSION

Data presented in Table 1 clearly showed significant influence of seed treatment on

Table 1. Effect of seed treatment on days to initiation of germination (days), germination percentage (%), germination rate index (%/days) and Germination Index of fishtail palm under Naturally Ventilated Poly-House at 90 DAP

Treatments	Initiation of germination (Days)		Germination Percentage (%)		Germination rate index (%/day)		Germination index	
	2022	2023	2022	2023	2022	2023	2022	2023
T ₁ Control (Open condition)	68.33	72.33	21.67	18.33	1.15	0.92	49.00	38.33
T ₂ NVPH + Control	62.33	56.00	38.33	38.33	2.08	2.15	89.33	91.67
T ₃ NVPH + sand paper	54.67	51.67	48.33	45.00	2.75	2.50	121.33	105.67
T ₄ NVPH + hot water	54.33	55.33	50.00	50.00	2.77	2.66	115.33	104.00
T ₅ NVPH + water	56.00	57.00	51.67	50.00	2.82	2.72	121.33	116.33
T ₆ NVPH + 50 % cow urine	61.33	56.67	50.00	50.00	2.74	2.74	117.67	116.67
T ₇ NVPH + 100 % cow urine	61.00	62.33	50.00	48.33	2.48	2.46	101.00	102.67
T ₈ NVPH + Coco peat container	65.00	66.33	48.33	45.00	2.38	2.19	95.67	84.00
T ₉ NVPH + Profenol 50 %	65.67	67.00	50.00	51.67	2.43	2.54	95.00	100.00
T ₁₀ NVPH + GA ₃ 1000 ppm	48.33	47.00	86.67	85.00	5.90	5.70	255.00	243.67
T ₁₁ NVPH + Bio NP	47.00	55.33	60.00	61.67	3.48	3.63	152.67	158.00
T ₁₂ NVPH + HNO ₃ 50%	46.33	50.33	68.33	68.33	4.43	4.47	194.67	197.67
CD (P=0.05)	5.67	6.07	9.21	7.69	0.46	0.39	21.41	20.37

Table 2. Effect of seed treatment on Coefficient of velocity of germination, Timson's Germination Index, Modified Timpson Germination Index and Survival Percentage of fishtail palm under Naturally Ventilated Poly-House at 90 DAP

Treatments	Coefficient of velocity of germination (%/Day)		Timson's Germination Index		Modified Timpson germination index (%)		Survival Percentage (%)	
	2022	2023	2022	2023	2022	2023	2022	2023
T ₁ Control	5.13	4.94	245	192	8.17	6.39	21.67	23.33
T ₂ NVPH + Control	5.18	5.25	447	458	14.89	15.28	43.33	45.00
T ₃ NVPH + sand paper	5.42	5.23	607	528	20.22	17.61	53.33	48.33
T ₄ NVPH + hot water	5.16	4.87	577	520	19.22	17.33	58.33	58.33
T ₅ NVPH + water	5.20	5.17	607	582	20.22	19.39	58.33	53.33
T ₆ NVPH + 50 % cow urine	5.19	5.18	588	583	19.61	19.44	53.33	58.33
T ₇ NVPH + 100 % cow urine	4.80	4.92	505	513	16.83	17.11	56.67	53.33
T ₈ NVPH + Coco peat container	4.74	4.60	478	420	15.94	14.00	58.33	60.00
T ₉ NVPH + Profenol 50 %	4.65	4.69	475	500	15.83	16.67	51.67	48.33
T ₁₀ NVPH + GA ₃ 1000 ppm	6.18	6.02	1275	1218	42.50	40.61	88.33	86.67
T ₁₁ NVPH + Bio NP	5.49	5.52	763	790	25.44	26.33	73.33	68.33
T ₁₂ NVPH + HNO ₃ 50%	5.99	6.07	973	988	32.44	32.94	83.33	76.67
CD (P=0.05)	0.48	0.50	107	102	5.67	6.07	5.44	9.21

initiation of germination (days), Germination percentage (%) Germination rate index (%/day) and germination index at 90 days after sowing. During first year, seeds treated with HNO₃ 50 % for 5 minutes recorded significantly lesser days to initiation of germination (46.33 days) while second year, treatment T₁₀ i.e. seeds treated with GA₃ 1000 ppm for one day recorded significantly lower days to initiation of germination (47.00 days). Moreover, seeds treated with GA₃ 1000 ppm for one day recorded significantly higher germination percentage (86.67% and 85.00%), Germination rate index (5.90%/day and 5.70%/day) and germination index (255 and 243.67) at 90 days after sowing during first and second years respectively. It was evident from the data depicted in Table 2 that germination treatment significantly influence Coefficient of velocity of germination, Timson's germination index, Modified Timpson germination index and survival percentage at the 90 days after sowing. Seeds treated with GA₃ 1000 ppm for one day recorded significantly higher Coefficient of velocity of germination (6.18 and 6.02), Timson's germination index (1275 and 1218), Modified Timpson germination index (42.50% and 40.61%) and survival percentage (88.33% and 86.67%) at the 90 days after sowing during first and second years respectively. Fishtail palm seeds without any treatment kept in open condition recorded significantly poor performance in all the germination parameters as compared to treatments kept under naturally ventilated poly-house along with lowest germination (21.67% and 18.33%) and survival percentage (21.67% and 23.33%) after 90 days after sowing during first and second years respectively.

GRI calculations merely show the percentage of germination per day for that treatment, so the higher the percentage and the shorter the duration, the higher the GRI. It has less information regarding the 'high' and 'low' germination days rather than it spreads the percentage evenly across the time spread. In case of CVG not focuses on the final germination percentage, but it emphasis on the time required for reaching it. While GI appears to be the most comprehensive observation tool combining both germination percentage and speed of germination (spread, duration and 'high or low' events) [17]. It also magnifies the variation among seed lots ultimately makes easier to compare numerical measurement and provide more insight to the germination behaviour. Amongst various treatments under study GA₃ shown better results comparing over other

treatments for germination and survival of seeds in fishtail palm might be due to its role and plant growth regulator and enhancing the seed elongation and amylase hydrolysis process which is activated by giberellic acid which allows radicle to push through the seed coat and ultimately breaking seed dormancy. The longer the seed is soaked with GA₃, the more GA₃ accumulates in the seed cell. Moreover, the increased germination percentage in GA₃ treated seeds might be attributed to fact that the GA₃ helps in breaking the seed dormancy which results in early and enhanced seed germination due to the diffusion of endogenous auxin and gibberellins like substances [18,19]. GA₃ might have helped in weakening of the tissues surrounding the embryo or the stimulation of embryonic elongation or both. Rout et al. [20] observed that increase in germination percentage with GA₃ might be due to involvement of GA₃ in the activation of cytological enzymes along with increase in cell wall plasticity and better water absorption. Provision of GA₃ as a growth regulator substance can stimulate cell division. The application of 1000 ppm GA₃ increased germination to palm kernels is thought be optimal in stimulating cell growth and ensuring that the kernels germinates fast. The results are in accordance with Setiawan et al. [21] in oil palm Kadir et al. [22] and Moussa et al. [1]. All the treated seeds recorded better germination percentage compared to control. With respect to germination percentage, seeds treated with Bio NP responded well among all the treatments after GA₃ and HNO₃ treatment respectively.

4. CONCLUSION

From the experiment it was evident that fishtail palm seed has very low germination percentage and slow rate of germination. It is must to provide a proper seed treatment get higher germination percentage along with uniform and quick germination of seed. The experiment does prove that seeds of fishtail palm treated with 1000 ppm GA₃ for one day resulted in lower initiation of germination along with higher germination speed, germination percentage and survival percentage. It was also clear that growing seeds under naturally ventilated poly house condition also help to improve germination. Nurseryman or farmer growing fishtail palm seeds are recommended to seed soaked in 1000 ppm GA₃ solution for one day and place them under naturally ventilated poly house condition in polybags for better germination and early growth.

5. FUTURE SCOPE

GA₃ gave promising results for breaking seed dormancy in fishtail palm and more research can be conducted to see the effect of different concentration along with duration to get the best results in improving initiation of germination. Moreover along with combination of other treatments can be tried to get better results in future.

CONSENT

As per international standards or university standards, respondents' written consent has been collected and preserved by the author(s).

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (Chat GPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Moussa H, Margolis HA, Dubé PA, Odongo J. Factors affecting the germination of doum palm (*Hyphaene thebaica* Mart.) seeds from the semi-arid zone of Niger, West Africa. *Forest Ecology and Management*. 1998;104(1-3):27-41.
2. Ehara H, Morita O, Komada C, Goto M. Effect of physical treatment and presence of the pericarp and sarcotesta on seed germination in sago palm (*Metroxylon sagu* Rottb.). *Seed Sci. Technol*. 2001;29:83–90.
3. Orozco-Segovia A, Batis AI, Rojas-Aréchiga M, Mendoza A. Seed biology of palms: A review. *Palms*. 2003;47:79–94.
4. Prakash K, Chaudhury R, Rohini MR, Singh B, Malik SK. Contrasting seed biology of two ornamental palms: Pygmy Date Palm (*Phoenix roebelenii* O'Brien) and Fishtail Palm (*Caryota urens* L.) and implications for their long-term conservation. *Indian Journal of Traditional Knowledge*. 2019;18(3):477-484.
5. Department of Export Agriculture. Annual report. Sri Lanka; 1995.
6. Kader (Al-Mударis) M. Notes on various parameters recording the speed of seed germination. *Journal of Agriculture in the Tropics and Subtropics*. 1998;99:147–154.
7. BRASIL. Ministério da Agricultura, Pecuária e Abastecimento. Regras para análise de sementes. Brasília: Secretaria Nacional de Defesa Agropecuária. 2009;399-400.
8. Scott S, Jones R, Williams W. Review of data analysis methods for seed germination. *Crop Science*. 1984;24:1192–1199.
9. Esechie H. Interaction of salinity and temperature on the germination of sorghum. *Journal of Agronomy and Crop Science*. 1994;172:194–199.
10. Goodchild NA, Walker MG. A method of measuring seed germination in physiological studies. *Annals of Botany*. 1971;35(141):615–621.
11. Bench AR, Fenner M, Edwards P. Changes in germinability, ABA content and ABA embryonic sensitivity in developing seeds of *Sorghum bicolor* (L.) Moench induced by water stress during grain filling. *New Phytologist*. 1991;118:339–347.
12. Labouriau LG. A germinação das sementes. Washington: Secretaria da OEA. 1983;173.
13. Jones K, Sanders D. The influence of soaking pepper seed in water or potassium salt solutions on germination at three temperatures. *Journal of Seed Technology*. 1987;11:97–102.
14. Timson J. New method of recording germination data. *Nature*. 1965;207(4993): 216.
15. Khan MA, Ungar IA. The effect of salinity and temperature on the germination of polymorphic seeds and growth of *Atriplex triangularis* Willd. *American Journal of Botany*. 1984;71(4):481--489.

16. Panse VG, Sukhatme PV. Statistical Methods for Agricultural Workers. Publication and Information Division, ICAR, New Delhi; 1967.
17. Kader MA. A comparison of seed germination calculation formulae and the associated interpretation of resulting data. Journal & Proceedings of the Royal Society of New South Wales. 2005;138: 65–75.
18. Gurung N, Swamy GSK, Sarkar SK, Ubale NB. Effect of chemicals and growth regulators on germination, vigour and growth of passion fruit (*Passiflora edulis* Sims.). The Bioscan. 2014;9(1):155-157.
19. Singh M, John SA, Rout S, Patra SS. Effect of GA₃ and NAA on growth and quality of garden Pea (*Pisum sativum* L.) cv. Arkel. The Bioscan. 2016;10(3):381-383.
20. Rout S, Beura S, Khare N, Patra SS, Nayak S. Effect of seed pre-treatment with different concentrations of gibberellic acid (GA₃) on seed germination and seedling growth of *Cassia fistula* L. Plants Stud. 2017;5(6):135-8.
21. Setiawan K, Irawati E, Ardian, Agustiansyah. The effect of difference soaking duration and GA₃ concentration on germination of oil palm (*Elaeis guineensis* Jacq). 2nd International Conference on Agriculture and Applied Science. 2021;1-7.
22. Kadir M, Image RC, Syatrawati, Nur AS. J. Agroplanta. 2020;9:95-104.

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