



The Effect of Air Dried Leaf Powder and Burnt Leaf Ash of Different Plants on Root-knot Nematode (*Meloidogyne* spp.) on Okra (*Abelmoschus esculentus* L. Moench)

O. F. Nnadi¹, C. C. Ononuju¹, E. A. Ikwunagu^{1*} and C. C. Orikara¹

¹Department of Plant Health Management, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. Author C. C. Ononuju designed the study and wrote the protocol. Author OFN carried out the experiment, managed the literature searches and wrote the first draft of the manuscript. Author C. C. Orikara carried out the laboratory part of the experiment while author EAI managed the analysis of the study and wrote the final manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JEAI/2019/v41i130389

Editor(s):

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Complete Peer review History: <https://sdiarticle4.com/review-history/49405>

Original Research Article

Received 12 April 2019
Accepted 26 June 2019
Published 02 October 2019

ABSTRACT

The effect of air dried leaf powder and burnt leaf ash of different plants on the control of root-knot nematode (*Meloidogyne* spp.) on Okra was investigated in a pot experiment. The experiment was laid out in a Completely Randomized Design with nine treatments each replicated four times. The treatments included: burnt leaf ashes and air dried leaf powders of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia*, a synthetic nematicide (Carbofuran 3G) and two control which included inoculated but untreated (control 1) and un-inoculated and untreated (control 2) pots. The plants were inoculated with 1000 nematode eggs (*Meloidogyne* spp.), two weeks after emergence. Three days after, the treatments were applied at the rate of 30 g each, while the synthetic

*Corresponding author: E-mail: emmanuelikwunagu@yahoo.com;

nematicide (Carbofuran 3G) at 3 g/per pot. Parameters recorded were: plant height, number of leaves, fresh and dry shoot weights, fresh root weight, and number and weight of pods per plant as plant growth parameters. Number of galls, number of nematode eggs in roots and number of juveniles in soil as nematode parameters. Results obtained indicated significant differences among the treatments in most parameters recorded and compared to controls. Generally, results from air dried leaf powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* were better and compared favorably with the nematicide treated plants.

Keywords: Air dried leaf; Burnt leaf ash; Carbofuran; *Meloidogyne* spp.; *Abelmoschus esculentus*; synthetic nematicide.

1. INTRODUCTION

Okra as a vegetable crop belongs to the genus *Abelmoschus*, Family *Malvaceae* and has two main species: *Abelmoschus esculentus* (L) Moench and *Abelmoschus caillei* (A. Chev) [1]. It is an annual crop mainly grown as a fruit and leafy vegetable in the tropics [2]. It is widely distributed in the Tropical and Subtropical Regions of Asia, Southern Europe, the Mediterranean countries and the Americas [3]. Nigeria is the second largest producer of Okra in the world after India and largest producer in Africa (5.8 million tonnes), followed by Cote d'Ivoire, Ghana and others [4]. In Nigeria, it is widely distributed and consumed in either fresh or dried forms [5].

Okra is important constituent of most local dishes in West Africa. It is used as a soup thickener and may also be served with rice and other food types [2]. In Nigeria, okra is grown across different ecological zones because it serves as a source of income to farmers as well as a cheap source of protein, vitamins A, B complex and C and mineral (Ca, P, Fe and K) to many households [6]. It is a nutritious vegetable that contains 86.1% water, 2.2% protein, 0.2% fat, 9.7% carbohydrates, 1.0% fiber and 0.8% ash [7], hence it plays vital role in human diet. Okra seeds contain greenish yellow edible oil which is also suitable for use as a bio-fuel [8].

Okra is notorious for its susceptibility to root knot nematodes [9]. *Meloidogyne* spp. are responsible for about 70-90% of the yield losses in okra [10]. Almost all the vegetables in tropical and warm temperate regions are severely attacked by plant-parasitic nematodes particularly root-knot nematodes (*Meloidogyne* spp.) [11]. *Meloidogyne* spp. cause wilting, chlorosis, stunted growth, formation of galls in roots often leading to destruction of roots, poor growth, yield and crop failure when nematode population exceeds economic threshold level [11,12]. The symptoms for identifying disease caused by

Meloidogyne spp. is the presence of galls, wilting, loss of vigor, yellowing of leaves [13]. They have caused up to 80% yield losses in heavily infested soils [14]. However the extent of damage is influenced by the cultivar, nematode species, level of soil infestation and environment [15]. It is therefore necessary to control root-knot nematodes in order to avoid or minimize yield losses in okra.

Management of root-knot nematodes with synthetic nematicide can be very effective [11]. However, there is public outcry against synthetic nematicides due to their toxicity, persistence and hazards posed to non-target species and agriculturists [16]. As a result there is a growing preference for plant products which are less harmful, effective, easily degraded, pollution free, leave no harmful residues, cheaper and not toxic to host plants and humans [17]. These plants if explored for nematicidal activity might be alternatives to synthetic nematicides [18].

The objective of this study is to determine the effect of air dried leaf powder and burnt leaf ash of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* on the population of root-knot nematode (*Meloidogyne* spp.) on the growth and yield of okra, gall formation and also to compare the effect of these plant materials with the synthetic nematicide (Carbofuran 3G).

2. MATERIALS AND METHODS

2.1 Location of Experimental Site

The experiment was carried out in the Department of Plant Health Management, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria located on latitude 5°2'N and longitude 7°33'E. It lies in the humid tropical rain forest zone with annual rainfall of 1916 mm per annum, altitude of 112 m above sea level and relative humidity of 76% with temperature range of 19-35°C [19].

2.2 Experimental Design

The experiment was laid out in a Completely Randomized Design (CRD) on a platform in an open field using plastic pots, with nine treatments replicated four times including the control, giving a total of 36 pots.

2.3 Experimental Materials

2.3.1 Source of seed

Seeds of okra variety *Pusa suwaan* were obtained from National Root Crop Research Institute Umudike, Abia State, Nigeria.

2.3.2 Treatments

The materials used were: burnt leaf ash and air dried leaf powder of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* and a synthetic nematicide (Carbofuran 3G).

2.3.3 Treatments' sources and preparation

The plants were obtained within the school premises, while the synthetic nematicide (Carbofuran 3G) was obtained from a chemical store. The leaves were collected and washed under running tap to remove soil particles and reduce contamination by microorganisms. They were then spread on a clean platform for air drying. This was done to reduce the moisture content after which each material was divided into two parts: one part was ground into powder using an electric blender while the other parts were burnt to ash in a controlled environment and each part were put in a clean plastic container ready for application.

2.3.4 Soil preparation and sterilization

Top soil was collected and sifted to remove large soil particles, stones and plant debris. The soil was poured into a cut metal drum. It was moistened as a source of heat applied underneath. It was sterilized at 80°C for 20 minutes after which it was left to cool [20].

2.3.5 Extraction of nematode eggs from inoculum

Eggs of root-knot nematodes were extracted from the heavily galled roots of *Basella alba* (Ceylon spinach). The galled roots were washed in tap water and cut into pieces of 2 cm long, placed in a beaker containing 200 mls of 0.5%

NaOCl (Sodium hypochloride) solution and shaken vigorously for about 5 minutes [21]. This was done to prevent egg damage, while the gelatinous matrix is being removed. The solution was poured over two nested sieves of 75 µm mesh 25 µm mesh to collect the eggs. Eggs in the 25 µm mesh sieve were rinsed with cold water and washed into a beaker. The cut roots in the original beaker were washed twice with water to obtain additional eggs. The number of eggs in 1 ml of water was estimated by counting four samples of a milliliter each using Doncaster's counting dish under a stereomicroscope and the average was taken [21].

2.3.6 Sowing of seeds

Okra seeds were sown at two seeds per hole in plastic pots with diameter 22.5 cm containing 6 kg sandy loam soil mixture that has been sterilized. Two weeks after, the seedlings were thinned down to a healthy plant per pot.

2.3.7 Inoculation of plants with nematode eggs

Three weeks after emergence, the plants were inoculated with a calculated volume of 1000 eggs of *Meloidogyne* spp. Eggs were applied near the base of plant by making holes around the Okra plants.

2.3.8 Application of treatments

Thirty (30 g) each of the air dried leaf powder and burnt leaf ash of *B. nitida*, *P. macrophylla* and *D. regia* were applied evenly on the surface of the soil in each bucket according to the treatments and their replications three days after inoculation. A synthetic nematicide (Carbofuran 3G) at 3.0 g ai per plant was applied. The controls include plants whose soils were inoculated but untreated (control 1) and uninoculated and untreated (control 2) respectively.

The treatment combinations were as follows:

1. T1 Nematode + Burnt leaf ash of *Baphia nitida*
2. T2 Nematode + Air dried leaf ash of *Baphia nitida*
3. T3 Nematode + Burnt leaf ash of *Pentaclethra macrophylla*
4. T4 Nematode + Air dried leaf ash of *Pentaclethra macrophylla*

5. T5 Nematode +Burnt leaf ash of *Delonix regia*
6. T6 Nematode + Air dried leaf ash of *Delonix regia*
7. T7 Nematode + synthetic nematicide (Carbofuran 3G).
8. T8 Inoculated but untreated (Control 1)
9. T9 Un-inoculated and untreated (Control 2)

(NPK 15:15:15) was applied at the rate of 100 kg/ha to give a rate of 0.4 g per bucket. Water was applied to the plant as required.

2.4 Data Collection

Data collected at the end of the experiment after twelve weeks included:

- Plant height- measured using a meter rule in centimeter.
- Number of leaves- counted visually.
- Fresh and dry shoot and fresh root weights- measured using a digital laboratory weighing balance in grams (g).
- Number of eggs in root and juveniles in soil- extracted using the modified Baermann technique [22], and counted using Domncaster's counting dish under an electronic stereomicroscope.
- Number of galls in roots- counted visually.
- Number of pods- counted visually.
- Weight of pods- measured in grams (g) using a digital laboratory weighing balance.

2.5 Statistical Analysis

The data collected were subjected to Analysis of Variance (ANOVA) and means were separated using Least Significant Difference (LSD) at 5% probability level ($P < 0.05$) by using computer software "Genstat Discovery Edition 4".

3. RESULTS AND DISCUSSION

The effect of treatments on plant height and number of leaves are shown in Table 1. As for plant height, significant differences ($P < 0.05$) were observed between control 2 (un-inoculated and untreated) (27.17 cm) and all the treatments being less than control 2. There was also a significant difference between the plants treated with air dried leaf powder of *B. nitida* (18.77 cm) and control 1 (6.65 cm). As for number of leaves,

there was no significant difference recorded among the treatments including the controls. It is worthy noticed that the highest number of leaves (15.00) was recorded for the plants treated with burnt leaf ash of *B. nitida*, compared to the lowest (6.00 cm) was recorded for control 1 (inoculated but untreated).

Table 2 shows the effect of treatments on number and weight of pods. There was no significant difference ($P < 0.05$) between the treatments and the controls on the number of pods. It is worthy noticed that the highest number of pods was recorded by control 2 (11) and equals to that recorded to plants treated with synthetic nematicide (11), with the least number of pods (5.0) recorded for (control 1). Significant differences ($P < 0.05$) were recorded on the weight of pods. All the treatments were significantly less from control 2 (34.9 g), except the plants treated with synthetic nematicide (28.3 g). Also, there were significant differences between all treatments and control 1.

The effect of treatments on fresh and dry shoot and fresh root weights was illustrated in Table 3. As for fresh shoot weight, air dried leaf powder of *B. nitida* (480 g) which differed significantly compared with control 1 (205 g) and burnt leaf ash of *B. nitida* (133 g) but did not with control 2 and other treatments. Regarding dry shoot weight, there was no significant difference recorded among the treatments including the controls. The highest dry shoot weight (85.2 g) was recorded for the plants treated with burnt leaf ash of *P. macrophylla* compared to the lowest (30 g) was recorded for control 1. Regarding fresh root weight all the treatments significantly differed from control 2 (371 g). However, the same treatments did not differ significantly from control 1 and from each other.

Effect of treatments on the number of nematode eggs and galls in roots, and juveniles in soil are shown in Table 4. As for number of galls in roots, all the treatments significantly reduced number of galls and compared favourably with control 1, but the treatments did not differ significantly from each other. Similar observations were made on the number of eggs in root. As for number of juveniles in soil, the treatments and the controls did not differ significantly from each other. However apart from control 2 (000), plants treated with nematicide had the least number of juveniles (126), while the highest number (475) was recorded for plants treated with burnt leaf ash of *P. macrophylla*.

Table 1. Effect of treatments on the plant height and number of leaves of okra infected with root-knot nematode (*Meloidogyne* spp.)

Treatments	Plant height (cm)	No. of leaves per plant
Burnt leaf ash of <i>Baphia nitida</i>	11.20	15.00
Air dried leaf powder of <i>Baphia nitida</i>	18.77	13.00
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	12.62	8.00
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	12.00	10.00
Burnt leaf ash of <i>Delonix regia</i>	11.07	13.00
Air dried leaf powder of <i>Delonix regia</i>	7.25	10.00
Synthetic nematicide (Carbofuran 3G)	10.25	12.00
Inoculated but untreated (control 1)	6.65	6.00
Un-inoculated and untreated (control 2)	27.17	11.00
LSD _(0.05)	6.25	NS

Where, NS = No Significant Difference

Table 2. Effect of treatments on the number and weight of Okra pods

Treatments	No. of pods per plant	Weight of pods (g)
Burnt leaf ash of <i>Baphia nitida</i>	7.0	21.3
Air dried leaf powder of <i>Baphia nitida</i>	10.0	23.4
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	7.0	15.6
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	8.0	21.4
Burnt leaf ash of <i>Delonix regia</i>	10.0	22.0
Air dried leaf powder of <i>Delonix regia</i>	6.0	16.1
Synthetic nematicide (Carbofuran 3G)	11.0	28.3
Inoculated but untreated (control 1)	5.0	6.1
Un-inoculated and untreated (control 2)	11.0	34.9
LSD _(0.05)	NS	9.75

Where, NS = No Significant Difference

Table 3. Effect of treatments on fresh shoot, dry shoot and fresh root weights of Okra infected with root-knot nematode (*Meloidogyne* spp.)

Treatments	Fresh shoot weight (g)	Dry shoot weight (g)	Fresh root weight (g)
Burnt leaf ash of <i>Baphia nitida</i>	133.0	32.8	66
Air dried leaf powder of <i>Baphia nitida</i>	480.0	76.8	187
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	335.0	85.2	118
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	326.0	46.2	57
Burnt leaf ash of <i>Delonix regia</i>	308.0	64.0	136
Air dried leaf powder of <i>Delonix regia</i>	389.0	63.5	165
Synthetic nematicide (Carbofuran 3G)	362.0	70.0	132
Inoculated but untreated (control 1)	205.0	30.0	122
Un-inoculated and untreated (control 2)	629.0	68.2	371
LSD _(0.05)	267.1	NS	133.2

Where, NS = No Significant Difference

The results on the effect of the treatments on plant height agreed with the findings of Bawa et al. [23] in which the plant extracts used had a significant effect on the height of tomato. They

are still in agreement with the findings of Kayani et al. [13] who confirmed that the symptoms for identifying diseases caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,

yellowing of leaves. Also Perry et al. [24] confirmed that the symptoms also include, yellowing of leaves and stunting, loss of vigor, wilting due to lack of moisture, decay of tissue due to secondary infection and yield loss. The growth reduction in crops such as Okra due to nematode infestation varies depending on its population density, level of cultivar susceptibility and environmental condition to which the host is subjected to [25].

The reduction in the fresh shoot and root weight could be due to slow rate of action of the active ingredients in the plant materials [26], or due to poor nutrients and water flow as a result of giant galls formed on the plant roots [27] which inhibit the normal growths and development of shoots and roots of plants affected.

The reduced number of nematode population and number of galls compared to control 1 agrees with the observations of Adegbite and Adesiyani [28] who indicated that as the higher the nematode population, the higher the number of galls and the higher the amount of damage occurred to plant which hinders the transportation of water and mineral nutrients from roots to aerial parts of the plant. The results obtained confirmed the findings of Ozores-Hampton [29] that the use of organic amendments suppressed soil phyto-parasitic nematode populations. Abulusoro et al. [30] reported that the susceptible tomato plants infected with root-knot nematodes (*Meloidogyne* spp.) show stunted growth, yield loss and conspicuous root galls, but that a number of plants are thought to contain biologically active ingredients which when applied in the soil reduce

the incidence of plant-parasitic nematodes. Sikora and Fernandez [11] also found that application of sesame seed extract reduced the incidence of root-knot nematodes and the severity of galling on okra roots. According to Akhtar and Mahmood [31], sesame seed extracts have a systemic activity against nematodes which may have accounted for the lower number of galls and lower mean population in treated plants. Also Vats et al. [32] reported reduction of galls and egg masses when some *M. javanica* infected tomato plants were treated with leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis*.

The reduced number and weight of pods agrees with Bolles and Johnson, [33] who reported that nematode (*Meloidogyne* spp.) greatly decreased and delayed the production of pods. Also, Adesiyani et al. [34] reported that some crop varieties could be effective hosts of root-knot nematode but suffer no statistically significant damage. However the extent of damage is greatly influenced by the cultivar, nematode species, level of soil infestation and environment [15]. According to Gommers [35] active compounds with nematocidal activity which have been found to limit the activities of plant-parasitic nematodes in plants includes alkaloids, diterpenes, fatty acids, glucosinolates, isothiocyanates, phenols, polyacetylenes, sesquiterpenes and thienyls. These compounds were extracted from different plants part such as roots, leaves and so on which could be responsible for the positive effects of air dried leaf ash and burnt leaf ash of these plants on the population of nematode in the soil and root as well as in pod formation.

Table 4. Effect of treatments on the population of *Meloidogyne* spp. in roots and soil

Treatments	No. of galls in roots	No. of eggs in roots	No. of juveniles in soil
Burnt leaf ash of <i>Baphia nitida</i>	0.75	126.00	350
Air dried leaf ash of <i>Baphia nitida</i>	0.25	102.00	250
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	0.50	102.00	475
Air dried leaf ash of <i>Pentaclethra macrophylla</i>	1.75	202.00	351
Burnt leaf ash of <i>Delonix regia</i>	1.00	158.00	425
Air dried leaf ash of <i>Delonix regia</i>	0.75	126.00	300
Synthetic nematicide (Carbofuran 3G)	0.00	78.00	126
Inoculated but untreated (control 1)	5.00	575.00	425
Un-inoculated and untreated (control 2)	0.00	000.00	000
LSD _(0.05)	2.23	272.5	NS

Where, NS = No Significant Difference

4. CONCLUSION AND RECOMMENDATION

The different treatments significantly decreased the population of root-knot nematode (*Meloidogyne* spp.). There were significant differences between the controls and the treatments in some of the parameters measured, while there were no significant differences in others. Synthetic nematicide (Carbofuran 3G) appeared to be more effective in the control of root-knot nematode. However, the performance of the synthetic nematicide was not significantly different from the results obtained from air dried powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* at the rate applied. From the findings of this study, air dried powder of *B. nitida* and burnt leaf ash of *D. regia* appeared to have significantly performed better than other treatments and is being recommended as an alternative for the synthetic nematicide (Carbofuran 3G) in the control of root-knot nematode. Notwithstanding, further research work both in green house and field trials at different rates are needed in order to authenticate this findings.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Siemonsma JS. The cultivation of Okra (*Abelmoschus* spp.), tropical fruit-vegetable (with special reference to the Ivory Coast) D.H.O. Thesis, Wageningen Agricultural, Wageningen, the Netherland. 1982;297.
- Tamiyu RA, Ahmed HG, Muhammad AS. Effect of sources of organic manure on growth and yield of okra (*Abelmoschus esculentus* (L.) Moench) in Sokoto, Nigeria. Nigerian Journal of Basic and Applied Science. 2012;20(3):213-216.
- Andres CD, Simandi B, Orsi F, Lambrou C, Tatla DM, Panayiotou C. Supercritical carbon dioxide extraction of okra (*Hibiscus esculentus* L.) seeds. Journal of the Science of Food and Agriculture. 2005;85: 1415–9.
- FAOSTAT. Food and Agricultural Organization of the United Nations: Okra Production Statistic; 2011. (Retrieved on 31 January, 2014) Available:<http://faostat.fao.org/foostat/>
- Farinde AJ, Owolarafe OK, Ogungbemi OI. An overview of production, processing, marketing and utilisation of okra in Egbedore Local Government Area of Osun State, Nigeria. Agricultural Engineering International: Manuscript. No. MES 07 002 the CIGR E-journal. 2007;9:1-17.
- Adebisi MA, Akintobi DC, Oyekale KO. Preservation of okra seed vigor by seed treatment and storage containers. Nigerian Journal of Horticultural Science (NJHS). 2007;12:1-7.
- Saifullah M, Rabbani MG. Evaluation and characterization of okra (*Abelmoschus esculentus* (L) Moench). Genotypes. J. Agric. 2009;7:92-99.
- Farroq A, Umer R, Muhammad A, Muhammad N. Okra (*Hibiscus esculentus*) seed oil for biodiesel production. Applied Energy. 2010;87(3):779-785.
- Noling JW. Nematode management in okra. Entomology and Nematology Department, Citrus Research Center, Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida, Lake Alfred, FL; 2012. Available:<http://edis.ifas.ufl.edu/> (Retrieved 23/07/2013)
- Saufiuddin S, Sheila S, Shweta S. Pathogenicity of root-knot nematode, *Meloidogyne incognita* and root-rot fungus, *Rhizoctonia solani* on okra (*Abelmoschus esculentus* L.). E-Journal of Science and Technology. 2011;3(6).
- Sikora RA, Fernández E. Nematode parasites of vegetables. In: Luc M, Sikora R.A, Bridge J, Eds. Plant Parasitic Nematodes in Subtropical and Tropical Agriculture. CAB International, Wallingford. 2005;319-390.
- Fourie H, McDonald AL, Loots GC. Plant parasitic nematodes in field crops in South Africa 6, Soybean. Nematology. 2001;3: 447-454.
- Kayani MZ, Mukhtar T, Hussian MA. Association of root-knot nematodes (*Meloidogyne* spp.) with cucumber in the Pothwar region of the Punjab province of Pakistan. Int. J. Biotech. 2012;a(1-2):23-29.
- Kaskavalci G. Effect of soil solarization and organic amendment treatment for controlling *Meloidogyne incognita* in tomato cultivars in Western Anatolia. Turkish. J. Agric. 2007;31:156-167.
- Ononuju CC. Management of plant parasitic nematode problems in commercial plantain and banana

- productions. A Paper Presented at the Third Annual Conference of Agricultural Society Institute Baddaggi, Nigeria; 1999.
16. Oka Y, Shuker S, Tkachi N, Trabelcy B, Gerchman Y. Nematicidal activity of *Ochradenus baccatus* against the root-knot nematode, *Meloidogyne javanica*. *Plant Pathology*. 2014;63(1):221-231.
 17. Amadioha AC. Evaluation of some plant leaf extracts against *Colletotrichum lindemuthianum* in cowpea. *Acta Phytopathologica et Entomologica Hungarica*. 2003;3(3-4):259-265.
 18. Siji JV, Jayaprakas CA, Sheela MS, Mohandas C. Efficacy of *Cleome viscosa* L. against *Meloidogyne incognita* infestation in okra (*Abelmoschus esculentus* L.) *Thai Journal of Agricultural Science*. 2010;43(3):151-156.
 19. N.R.C.R.I. National Root Crops Research Institute. NRCRI, Umudike, Meteorological Station Data; 2010.
 20. Ononuju CC, Ikwunagu EA, Okorocho AD, Okorie CC. Effects of different agricultural wastes and botanical on root knot nematode (*Meloidogyne* spp) on okra (*Abelmoschus esculentus* L. Moench). *Journal of Entomology and Nematology*. 2014;6(5):56-61.
 21. Hussey RS, Barker KR. A comparison of methods of collecting inoculum of *Meloidogyne* spp. including a new technique. *Plant Disease Reporter*. 1973;57:1025-1028.
 22. Hooper DJ. Extraction and handling of plant and soil nematode. *Nematodes of Tropical Crops*. 1969;21-22.
 23. Bawa JA, Mohammed I, Liadi S. Nematicidal effect of some plant extracts on root-knot nematodes (*Meloidogyne incognita*) of tomato (*Lycopersicon esculentum*). *World Journal Life Sci. and Medical Res*. 2014;3(3):4-7.
 24. Perry RN, Moens M, Starr JL. Root-knot nematodes. Wallingford, UK, CAB International; 2009.
 25. Ononuju CC, Fawole B. Evaluation of Ethoprophos Isazophos, Carbofuran for the control of plant parasitic nematodes and their effects on plantain growth and yield. *Nigerian J. Sci*. 2000;34:81-89.
 26. Barman M, Das P. Effect of chemical seed dressing and organic amendments alone and in combination for the management of root-knot nematode on green gram. *Indian J. Nematol*. 1996;26:72-76.
 27. Ploeg A. When nematodes attack is important. *California Grower*. 2001;12-13.
 28. Adebite AA, Adesiyon SO. Root extracts of plants to control root-knot nematode on edible soyabean. *World Journal Agricultural Science*. 2005;1:18-21.
 29. Ozores-Hampton M. Organic materials in horticulture: An industry perspective. *Intro. Hor. Tech*. 2002;12(3):8-9.
 30. Abulusoro SA, Oyedunmade EA, Olabiya TI. Screen house and laboratory assessment of toxic effect of brimstone (*Morinda lucida*) leaf of the rook-knot nematode, *Meloidogyne incognita*. *Plant Pathology Journal*. 2004;1:45-49.
 31. Akhtar M, Mahmood I. Control of plant parasitic nematodes with "Nimin" and some plant oils by bare-root dip treatment. *Nematologia Mediterranean*. 1993;21:89-92.
 32. Vats R, Nandal SN, Dalal MR. Comparative efficacy of different preparations of leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis* against *M. javanica* in tomato nursery. *Haryana Agricultural University Journal of Research*. 1996;26(2):99-102.
 33. Bolles B, Johnson L. Nematode damage of Okra. University of Florida. IFAS extension; 2012. Available:<http://Washington.ifas.ufl.edu/> (Retrieved 23/07/2013)
 34. Adesiyon SO, Caveness FE, Adeniji MO, Fawole B. The root-knot nematodes, *Meloidogyne* spp. In *Nematode Pests of Tropical Crops*. Heinemann Books (Nigeria) Ltd. 1990;Chapter 3:19-34.
 35. Gommers FJ. Biochemical interaction between nematodes and plants and their relevance to control. *Helminthological Abstract Series B Plant Nematology*. 1981;50:9-24.

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